

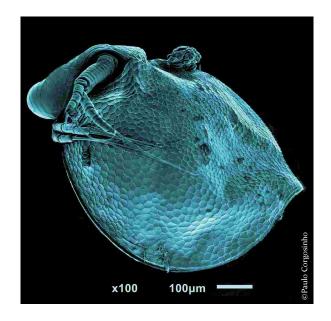
RE:	Toxicity Tests of Effluent from the Y-12 National Security Complex Outfall 200 Conducted May 3-10, 2023
From:	L.M. Stevenson, 1504, MS-6351 (865-341-0398).
c:	S. Loveless, J. Stinnett, K. Kinder, T.J. Mathews, P. Ku, A.M. Fortner
То:	K.G. Hanzelka (RC)
Date:	May 23, 2023

Appended are the results of toxicity tests of effluent from Outfall 200 conducted from May 3 to May 10, 2023. The effluent was evaluated for toxicity with fathead minnows (*Pimephales promelas*) and water fleas (*Ceriodaphnia dubia*). For both species, the Inhibition Concentration₂₅ (IC₂₅) for survival, growth, and/or reproduction for organisms exposed to effluent from Outfall 200 was >100% (the highest concentration of effluent tested). The NPDES permit states that toxicity will be demonstrated if the IC₂₅ is less than or equal to the permit limit (50% effluent for Outfall 200). All of the results for all endpoints were within permit limits.

Outfall	Test Organism	Endpoint	IC ₂₅
Outfall	Fathead	Survival	>100%
200	minnow	Growth	>100%
Outfall	Ceriodaphnia	Survival	>100%
200		Reproduction	>100%

Please do not hesitate to call if you have any questions or comments.

Attachment lms



Ceriodaphnia dubia TOXICITY TEST REPORT

Test Number 2985 | Y-12 National Security Complex Outfall 200 | 17 May 2023

Toxicology Laboratory Principal Investigator: Dr. Louise Stevenson Environmental Sciences Division Oak Ridge National Laboratory Building 1504 P.O. Box 2008, MS 6351 Oak Ridge, TN 37831-6351 (865) 341-0398

STANDARD REPORT FORM CERIODAPHNIA 3-BROOD SURVIVAL AND REPRODUCTION TEST

Test Number 2985 | Start Date: 3 May 2023 | End Date: 10 May 2023

1. INTRODUCTION

- 1.1 Permit Number: TN0002968
- 1.2 Toxicity testing requirements of permit: A 3-brood *Ceriodaphnia* Survival and Reproduction Test and a 7-day Fathead Minnow (*Pimephales promelas*) Larval Survival and Growth Test shall be conducted annually. All tests will be conducted using a minimum of three 24-hour composite samples of final effluent. The measured endpoint for toxicity will be the inhibition concentration causing 25% reduction (IC₂₅) in survival, reproduction, or growth of the test organisms as compared to the controls.

The permit states that toxicity is demonstrated if the IC₂₅ is less than or equal to the permit limit. The permit limit for Outfall 200 is 50% whole effluent.

- 1.3 Plant location: Y-12 National Security Complex.
- 1.4 Name of receiving water body: East Fork Poplar Creek.

1.5 Contractor: Toxicology Laboratory Environmental Sciences Division Oak Ridge National Laboratory P.O. Box 2008, MS 6351 Oak Ridge, TN 37831-6351 (865) 576-3459

2. SAMPLE

- 2.1 Sample description: Effluent from Outfall 200.
- 2.2 Sampling point: NPDES Outfall 200.
- 2.3 Sampling period: 2 May 2023 to 8 May 2023
- 2.4 Sampling method: Three 24-h flow-proportionate composite samples of final effluent.
- 2.5 Samples were used immediately then stored at 4 ± 2 °C to be used for two or three days during the daily effluent renewal process.

- 2.6 Sample pre-treatment: Sample temperature was raised to 25 ± 1 °C in a warm water bath prior to test initiation and daily test renewal.
- 2.7 Sample information:

Parameter	Sample 1	Sample 2	Sample 3
Collection Start Date	5/2/2023	5/4/2023	5/7/2023
Composite Duration	24 h	24 h	24 h
Date of Delivery to ESD Tox Lab	5/3/2023	5/5/2023	5/8/2023
Chain-of-Custody Form Number	031101	031102	031103
Sample Temperature (°C)	14.3	11.6	14.7
рН (S.U.)	8.16	8.13	8.24
Conductivity (µS/cm)	544	525	418
Alkalinity (mg/L as CaCO ₃)	124	140	132
Hardness (mg/L as CaCO ₃)	290	250	200
Chlorine (Free/Total) (mg/L)	0.02/0.01	0.01/0.01	0.01/0.02

3. TEST ORGANISMS

- 3.1 Species: Ceriodaphnia dubia.
- 3.2 Life stage: Neonates ≤24 h old; all born within 8 h of each other.
- 3.3 Source: Environmental Sciences Division cultures.
- 3.4 Incubation water for cultures: 25% DMW [2.5:7.5 (v:v) ratio of degassed mineral water to deionized distilled water augmented with trace metals].
- 3.5 Temperature of cultures: 25 ± 1 °C.

4. TEST METHODS

- 4.1 Toxicity test method: Ceriodaphnia survival and reproduction test. Reference: EPA Test Method 1002.0, in P.A. Lewis et al., Short-term Methods for Estimating the Chronic Toxicity of Effluent and Receiving Waters to Freshwater Organisms, EPA/821/R/02/013 (4th Ed., October 2002; or most recent version).
- 4.2 End points of test: Survival and reproduction.
- 4.3 Modifications or deviations to Method 1002.0: None.

- 4.4 Date and time test started: 5/3/2023, 18:03
- 4.5 Date and time test terminated: 5/10/2023, 11:50
- 4.6 Type and volume of test chambers: Polystyrene microbeakers, minimum 15mL each.
- 4.7 Number of *Ceriodaphnia* per test chamber: 1.
- 4.8 Number of replicates per treatment: 10.
- 4.9 Dilution/control water: 25% DMW [2.5:7.5 (v:v) ratio of degassed mineral water to deionized distilled water augmented with trace metals].
- 4.10 Renewal period: 24 h
- 4.11 Test temperature: Mean = 25.2 °C; range = 25.0-25.3 °C.
- 4.12 Treatment groups/concentrations: Control, 12.5%, 25%, 50%, 75% and 100% of full-strength effluent.
- 4.13 Feeding regime during test: 100 μL of yeast-Cerophyl-trout food (YCT) mixture and 3 x 10⁶ cells of the green alga *Raphidocelis subcapitata* per 15 mL of test solution every 24 h from an algal stock with a concentration 3.0 3.5 x 10⁷ cells/mL (EPA/821/R/02/013; 4th Ed., October 2002; or most recent version).

5. QUALITY ASSURANCE

- 5.1 Standard toxicant used: Sodium chloride (source: Fisher Scientific).
- 5.2 Date of most recent chronic reference toxicant test: 04/19/2023 04/26/2023.
- 5.3 Dilution water used: 25% DMW [2.5:7.5 (v:v) ratio of degassed mineral water to deionized distilled water augmented with trace metals].
- 5.4 Survival IC₂₅ = 2.24 g NaCl/L; 95% C.I. = 1.74-2.32 g NaCl/L. Reproduction IC₂₅ = 1.38 g NaCl/L; 95% C.I. = 1.18-1.57 g NaCl/L. The IC₂₅s were calculated by the EPA linear interpolation method.
- 5.5 We report the most recent 20 tests, as recommended by EPA Chronic test guidelines (EPA, 2002).

Central tendency of IC₂₅ for survival: 1.49 ± 0.851 g NaCl/L (mean ± 2 SD).

CV of IC₂₅ for survival: 0.285 g NaCl/L

Central tendency of IC₂₅ for reproduction: 1.063 ± 0.619 g NaCl/L (mean ± 2 SD).

CV of IC₂₅ for reproduction: 0.291 g NaCI/L

A copy of the control chart is appended.

6. CERIODAPHNIA TEST RESULTS

Copies of the toxicity test logsheets are appended.

6.1 Summary of results from the *Ceriodaphnia* toxicity test:

Effluent Concentration	Number of replicates	Number of animals surviving for 3 broods	Mean number of offspring per female (±SD)
Control	10	10	30.7 ± 7.3
12.5%	10	10	20.7 ± 10.7
25%	10	10	24.7 ± 10.5
50%	10	10	26.9 ± 11.3
75%	10	10	30 ± 7.5
100%	10	10	28.9 ± 10.2

7. STATISTICAL ANALYSES

7.1 Survival

The calculated IC₂₅ for survival was >100% effluent.

7.2 Reproduction

The calculated IC₂₅ for reproduction was >100% effluent.

7.3 Summary of Ceriodaphnia toxicity test results:

IC₂₅ for survival: >100%

IC₂₅ for reproduction: >100%

8. SUMMARY OF CHEMICAL ANALYSES

8.1 Water quality of control water:

Parameter	Sample 1	Sample 2	Sample 3
pH (S.U.)	8.21	8.23	8.04
Conductivity (µS/cm)	241	235	246
Alkalinity (mg/L as CaCO₃)	80	80	120
Hardness (mg/L as CaCO₃)	130	130	130

8.2 Physical and chemical methods

pH, conductivity, and dissolved oxygen were measured using a YSI MultiLab 4010-3W.

The pH was measured by EPA method 150.1 with a YSI 4130 pH meter. The meter was calibrated with pH 4.0, 7.0, and 10.0 buffers.

Conductivity (µS/cm) was measured by EPA method 120.1 with a YSI 4310 meter. The meters were verified using certified reference standards.

Dissolved oxygen (mg/L) was measured by EPA method 360.1 with a YSI 4410W dissolved oxygen meter. The meter was calibrated in accordance with the manufacturer's instructions.

Alkalinity, hardness, and chlorine were measured using a Hach SL1000 Portable Parallel Colorimeter.

Instruments were calibrated and standardized according to manufacturer's instructions.

All measurements were made on fresh samples before daily water replacement. In addition, dissolved oxygen and pH were measured on water collected after daily replenishment period.

Report prepared by: Peijia Ku Report reviewed by: Louise Stevenson Date: 17 May 2023

Date: 22 May 2023 Louise Hevenson



Fathead Minnow TOXICITY TEST REPORT

Test Number 1690 | Y-12 National Security Complex Outfall 200 | 17 May 2023

Toxicology Laboratory Principal Investigator: Dr. Louise Stevenson Environmental Sciences Division Oak Ridge National Laboratory Building 1504 P.O. Box 2008, MS 6351 Oak Ridge, TN 37831-6351 (865) 341-0398

STANDARD REPORT FORM FATHEAD MINNOW SURVIVAL AND GROWTH TEST

Test Number 1692 | Start Date: 3 May 2023 | End Date: 10 May 2023

1. INTRODUCTION

- 1.1 Permit Number: TN0002968
- 1.2 Toxicity testing requirements of permit: A 3-brood *Ceriodaphnia* Survival and Reproduction Test and a 7-day Fathead Minnow (*Pimephales promelas*) Larval Survival and Growth Test shall be conducted annually. All tests will be conducted using a minimum of three 24-hour composite samples of final effluent. The measured endpoint for toxicity will be the inhibition concentration causing 25% reduction (IC₂₅) in survival, reproduction, or growth of the test organisms as compared to the controls.

Toxicity will be demonstrated if the IC_{25} is less than or equal to the permit limit. The permit limit for Outfall 200 is 50% whole effluent.

- 1.3 Plant location: Y-12 National Security Complex.
- 1.4 Name of receiving water body: East Fork Poplar Creek.
- 1.5 Contractor: Toxicology Laboratory Environmental Sciences Division Oak Ridge National Laboratory P.O. Box 2008, MS 6351 Oak Ridge, TN 37831-6351 (865) 576-3459

2. SAMPLE

- 2.1 Sample description: Effluent from Outfall 200.
- 2.2 Sampling point: NPDES Outfall 200.
- 2.3 Sampling period: 2 May 2023 to 8 May 2023
- 2.4 Sampling method: Three 24-h flow-proportionate composite samples of final effluent.
- 2.5 Samples were used immediately then stored at 4 ± 2 °C to be used for two or three days during the daily effluent renewal process. Samples were used within sample holding time guidance outlined in EPA Test Method 1000.

- 2.6 Sample pre-treatment: Sample temperature was raised to 25 ± 1 °C in a warm water bath prior to test initiation and daily test renewal.
- 2.7 Sample information:

Parameter	Sample 1	Sample 2	Sample 3
Collection Start Date	5/2/2023	5/4/2023	5/7/2023
Composite Duration	24 h	24 h	24 h
Date of Delivery to ESD Tox Lab	5/3/2023	5/5/2023	5/8/2023
Chain-of-Custody Form Number	031101	031102	031103
Sample Temperature (°C)	14.3	11.6	14.7
рН (S.U.)	8.16	8.13	8.24
Conductivity (µS/cm)	544	525	418
Alkalinity (mg/L as CaCO₃)	124	140	132
Hardness (mg/L as CaCO ₃)	290	250	200
Chlorine (Free/Total) (mg/L)	0.02/0.01	0.01/0.01	0.01/0.02

3. TEST ORGANISMS

- 3.1 Species: Fathead minnow (Pimephales promelas).
- 3.2 Hatch date: 1 May 2023 .
- 3.3 Life stage: Newly hatched larvae less than 48 h old.
- 3.4 Incubation water: Dechlorinated tap water.
- 3.5 Incubation temperature: 25 ± 1 °C.
- 3.6 Source: Cultures from Aquatic BioSystems, Inc., Fort Collins, CO.
- 3.7 Mean dry weight at test initiation: 0.129 mg
- 3.8 Diseases and treatment: None.

4. TEST METHODS

4.1 Toxicity test method: Fathead minnow larval survival and growth test. Reference: EPA Test Method 1000.0, in P.A. Lewis et al., *Short-term Methods for Estimating the Chronic Toxicity of Effluent and Receiving Waters to Freshwater Organisms*, EPA/821/R/02/013 (4th Ed., October 2002; or most recent version).

- 4.2 End points of test: Survival and growth.
- 4.3 Modifications or deviations to Method 1000.0: None.
- 4.4 Date and time test started: 5/3/2023, 16:30
- 4.5 Date and time test terminated: 5/10/2023, 16:30
- 4.6 Type and volume of test chambers: 600-mL borosilicate beakers, minimum 250 mL each.
- 4.7 Number of organisms per test chamber: 10.
- 4.8 Number of replicates per treatment: 4.
- 4.9 Dilution/control water: 25% DMW [2.5:7.5 (v:v) ratio of degassed mineral water to deionized distilled water augmented with trace metals].
- 4.10 Renewal period: 24 h
- 4.11 Acclimation of test organisms: Received larvae on 2 May 2023 at 13.5 °C.
- 4.12 Test temperature: Mean = 25.6 °C; range = 24.5-25.9 °C.
- 4.13 Treatment groups/concentrations: Control, 12.5%, 25%, 50%, 75% and 100% of full-strength effluent.
- 4.14 Feeding regime during test: Brine shrimp (*Artemia*) nauplii less than 24 h old; fed 1500 ± 100 shrimp per beaker twice daily.

5. QUALITY ASSURANCE

- 5.1 Standard toxicant used: Potassium chloride (source: Fisher Scientific).
- 5.2 Date of most recent chronic reference toxicant test: 05/03/2023 05/10/2023.
- 5.3 Dilution water used: 25% DMW [2.5:7.5 (v:v) ratio of degassed mineral water to deionized distilled water augmented with trace metals].
- 5.4 Survival IC₂₅ = 1.01 g KCI/L; 95% C.I. = 0.68-1.04 g KCI/L.

Growth IC₂₅ = 1.01 g KCI/L; 95% C.I. = 0.72-1.04 g KCI/L.

The IC₂₅s were calculated by the EPA linear interpolation method.

5.5 We report the most recent 20 tests, as recommended by EPA Chronic test guidelines (EPA, 2002).

Central tendency of IC₂₅ for survival: 0.868 ± 0.27 g KCI/L (mean ± 2 SD).

CV of IC $_{25}$ for survival: 0.156 g KCI/L

Central tendency of IC₂₅ for growth: 0.920 ± 0.228 g KCI/L (mean ± 2 SD).

- CV of IC₂₅ for growth: 0.125 g KCl/L
- A copy of the control chart is appended.

6. FATHEAD MINNOW TEST RESULTS

Copies of the toxicity test logsheets are appended.

6.1 Summary of results from the fathead minnow toxicity test:

	i				
Concentration	1	2	3	4	Mean
Control	1	1	1	1	1
12.5%	1	1	0.9	1	0.98
25%	1	1	0.8	1	0.95
50%	0.9	1	1	1	0.98
75%	1	0.9	1	1	0.98
100%	1	1	1	1	1

Survival

Dry Weight

Concentration	1	2	3	4	Mean ± SD
Control	0.82	0.78	0.75	0.75	0.78 ± 0.03
12.5%	0.86	0.67	0.69	0.79	0.75 ± 0.09
25%	0.74	0.87	0.71	0.77	0.77 ± 0.07
50%	0.73	0.85	0.91	0.7	0.8 ± 0.1
75%	0.75	0.7	0.72	0.82	0.75 ± 0.05
100%	0.71	0.69	0.82	0.78	0.75 ± 0.06

7. STATISTICAL ANALYSES

7.1 Survival

The calculated IC₂₅ for survival was >100% effluent.

7.2 Growth

The calculated IC₂₅ for growth was >100% effluent.

7.3 Summary of fathead minnow toxicity test results:

IC₂₅ for survival: >100%

IC₂₅ for growth: >100%

8. SUMMARY OF CHEMICAL ANALYSES

8.1 Water quality of control water:

Parameter	Sample 1	Sample 2	Sample 3
рН (S.U.)	8.21	8.23	8.04
Conductivity (µS/cm)	241	235	246
Alkalinity (mg/L as CaCO₃)	80	80	120
Hardness (mg/L as CaCO₃)	130	130	130

8.2 Physical and chemical methods

pH, conductivity, and dissolved oxygen were measured using a YSI MultiLab 4010-3W.

The pH was measured by EPA method 150.1 with a YSI 4130 pH meter. The meter was calibrated with pH 4.0, 7.0, and 10.0 buffers.

Conductivity (μ S/cm) was measured by EPA method 120.1 with a YSI 4310 meter. All values were corrected to 25°C. The meters were verified using certified reference standards.

Dissolved oxygen (mg/L) was measured by EPA method 360.1 with a YSI 4410W dissolved oxygen meter. The meter was calibrated in accordance with the manufacturer's instructions.

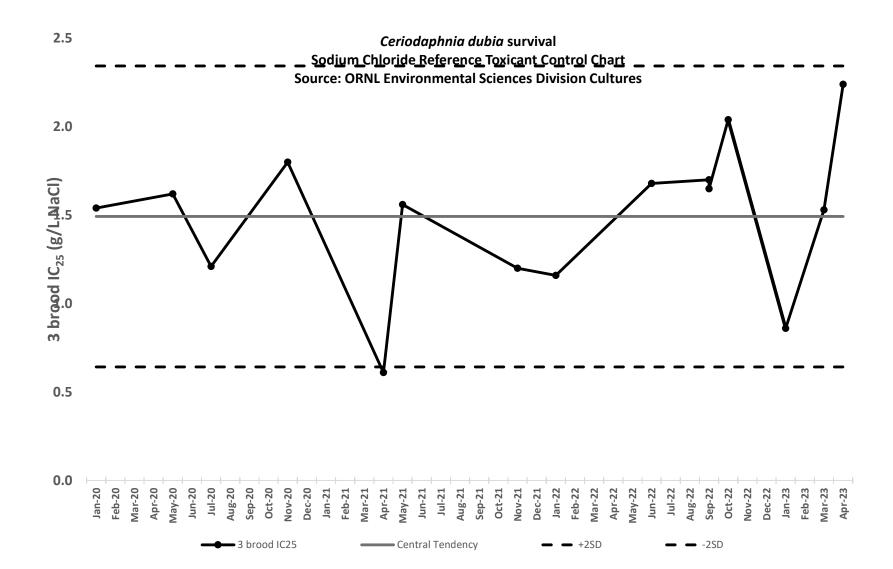
Alkalinity, hardness, and chlorine were measured using a Hach SL1000 Portable Parallel Colorimeter.

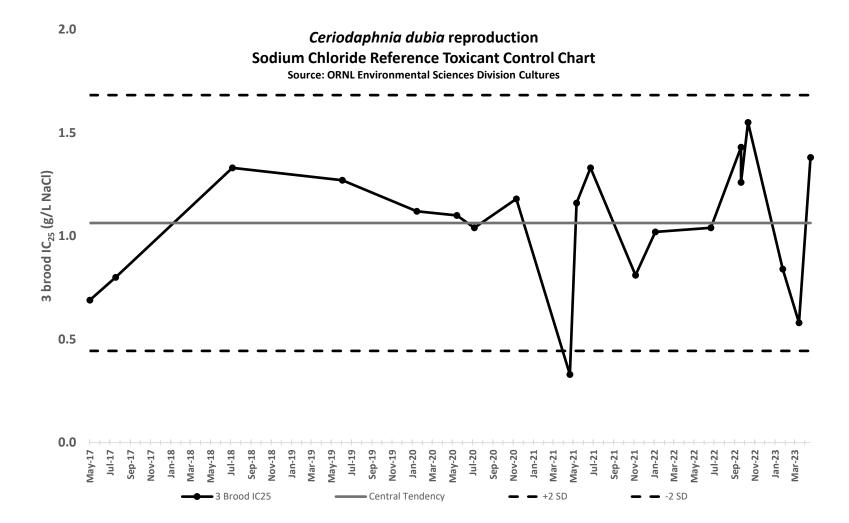
Instruments were calibrated and standardized according to manufacturer's instructions.

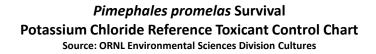
All measurements were made on fresh samples before daily water replacement. In addition, dissolved oxygen and pH were measured on water collected after daily replenishment period.

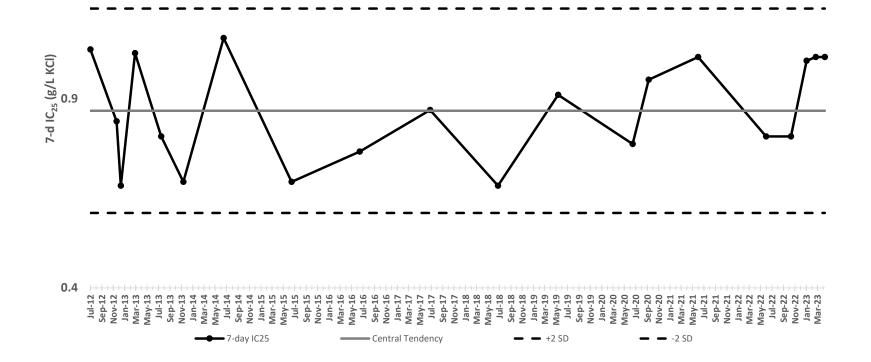
Report prepared by: Peijia Ku	Date: 18 May 2023		
Report reviewed by: Louise Stevenson	Date: 22 May 2023	louise	Stevensor

REFERENCE TOXICANT CONTROL CHARTS

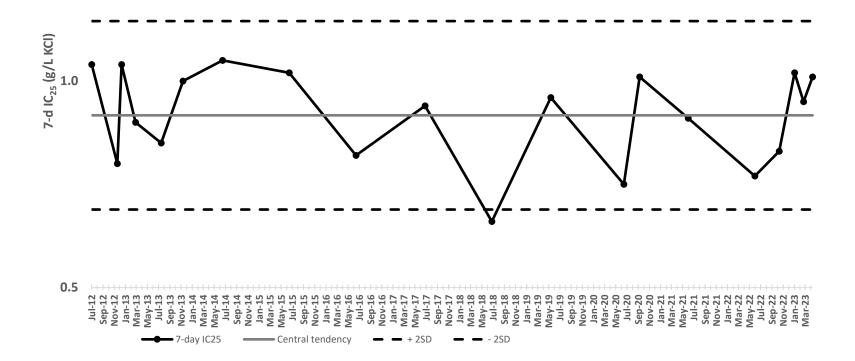








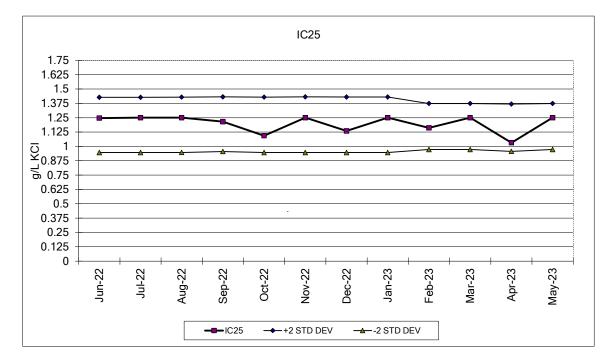
Pimephales promelas Growth Potassium Chloride Reference Toxicant Control Chart Source: ORNL Environmental Sciences Division Cultures





1300 Blue Spruce Drive, Suite C Fort Collins, Colorado 80524 Toll Free: 800/331-5916 Tel:970/484-5091 Fax:970/484-2514

Pimephales promelas



Chronic 7 Day Survival Test Data

IC 25 for Growth Test

L	LOEC	95% Confidence Avg. IC25	، ز	+2 STD	-2 STD
g/	(g/L KCI)	(upper) (lower) g/L KCl		DEV	DEV
	1.0	1.319 0.164 1.188		1.431	0.946
	1.0	1.250 1.144 1.188		1.431	0.946
	1.0	1.303 -0.506 1.173		1.374	0.972
	1.0	1.250 1.210 1.173		1.374	0.972
	1.0	1.272 0.023 1.163		1.369	0.957
	1.0	1.250 1.141 1.173		1.374	0.973
	1.0		1.250 1.141 1.173	1.250 1.141 1.173	1.250 1.141 1.173 1.374

**Current Test Dates: 5/3-10/2023

Aquatic BioSystems, Inc • Quality Research Organisms

WATER CHEMISTRY DATA LOGSHEETS

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7	1.5	Hardness (mg/L)	130				130		/	99999999999999999999999999999999999999
		pH (S.U.) Initial	8.207	8.19	8-23	8.20	8.04	8.20	8.05	219900000000000000000000000000000000000
		Final CD/FHM		8.59/8.05			8.541 7.93	8.55 7.90	8.57 7.91	8.28 8.10
	Control:	DO (mg/L) Initial	8.63	8.63	219	Pho	0 60			
		Final CD/FHM		8.71/7.76	9.09/6.97	8-78/7.15	8.79 [7.10	850 5.99	8.79 6.48	8.47 /7.43
		Conductivity (µS/cm)	277	277	271	261	280	261	251	
		Alkalinity (mg/L)						-/		
(\mathcal{D})		Hardness (mg/L)	/		/	/	/	/	/	
		Chlorine (mg/L) pH (S.U.) Initial	8.19	8.19	8.11	8.23	8.25	8.18	8.05	11111111111111111111111111111111111111
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	3	DO (mg/L) Initial	8.63	8.69	8.70	8.70	8.45	8.56	8.58	0.0)
		Final CD/FHM		8.6817.39	9.16/ 6.97		8.90/7.05	8.86 6.36		851/7.44
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(m)	0	Alkalinity (mg/L)		/		/		/	/	
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	25%	Final CD/FHM	8.77	8.51 8.08	8.60/8.12	8.61 / 8.10			8.63 7.98 8.74	8.36 8.04
		DO (mg/L) Initial	6.71	8.76	8.72 8.96/7.30	8.76	8.54 8.98 [7.4]	8.36	9.03/7	852/750
	_	Final CD/FHM Conductivity (µS/cm)	329	8.70/7.45 396	381	375	382	327	322 7.17	1.00
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×		Chlorine (mg/L)				/				
		pH (S.U.) Initial	8.16	8.19	8.14	8.18	8.22	8.25	8.17	
	8	Final CD/FHM		8.55/8.14	8.60/8.15	8.59/8.17		8.65/8.05	8.67 8.01	8.39 18.07
	50	DO (mg/L) Initial	8.76	8.77	8.87	8.78	8.58	8-74	8.95	a city of the
		Final CD/FHM				8.96/ 7.18	9.17 / 7.27		9.14 6.81	8.5417.49
0	I 8	Conductivity (µS/cm)	470	472	451	450	453	371	368	
C		Alkalinity (mg/L)						-/		12111212120000000000000000000000000000
		Hardness (mg/L)	/	/		/	/	/	/	MARINA
		Chlorine (mg/L) pH (S.U.) Initial	8.13	8.19	8.14	8.18	8.22	8.24	8.15	
	F.	Final CD/FHM	and the second real for an area in the second real for the	8.59/8.20	8.66/019	8.63/8.22		8.66 8.08	8.68 8.04	8.42/8.05
	1	DO (mg/L) Initial	9.02	8.91	8.99	8.90	8.61	860	9.02	
		Final CD/FHM		8-71/7.47	9.11/7.11	9.09/ 7.19		9.02 6.63	9.18 6.42	8.5317.36
		Conductivity (µS/cm)	344	549	525	520	525	418	415	11111111111111111111111111111111111111
0		Alkalinity (mg/L)	124	1	140			132	/	(<u>81118887898</u>)
(P)	L	Hardness (mg/L)	290		250			200	/	
		Chlorine (mg/L)	0,02/0.01	/	0.01/0.01			501/0.02	0 .11	01110200000000000000000000000000000000
	0	pH (S.U.) Initial	Contraction and the second second second second	8.19	8.13	8.18	8.21	8.24	8.14	8.44 8.05
5	10	Final CD/FHM		8.64 /8.24	8.67/8.25	8.67/821		8.5 8 24/8.1 9.58	9,20	THE PARTY
		DO (mg/L) Initial	4.12	8.91	9.11	8.83	8.64 9.25 7.00	9.09 6.20	820 6.69	8.58 7.3
	Env	Final CD/FHM		8.78/7.31	9.17 / 6.87	1.15/1.20	1.01 1.00	1 1 0.00	Rev. 04 20	
	Env	ironmental Sciences D	IVISION						101.0.20	

CHAIN OF CUSTODY FORMS

ENVIRONMENTAL SCIENCES DIVISION TOXICOLOGY LABORATORY CHAIN-OF-CUSTODY

DATE (MM/DD/YY) 05/03/23	ESD TEST NAME	OX TEST	NAME OF SAMPLER	s GARLAND / U	KIILIAMS	FRIG	CHAIN-OF-CUSTODY NO. 031101
SAMPLE NAME	OUTFALL NUMBER	SAMPLING TIME	SAMPLE TYPE *	NO. OF CONTAINERS	TOTAL VOLUME	ТЕМР (°С)	# 7009 TEMP. C/2
TOX TEST	200	1240	C	/	-218L	4°	14.3° <0.05
							7624
					n		
-					1		
THERMOMETER NO.	and a second	s	-				
SAMPLES RELINQUISHED BY	. S. Dar	Paul		-	DA	5/3/2	3 1308 DAM
SAMPLES RECEIVED BY	zijia Ku				DA	TE 5/3/23	
UCN-18631 (3 3-92)			RAB (G), 24 HR	COMPOSITE (C), OR	OTHER (O; DESCRIBE)		

ENVIRONMENTAL SCIENCES DIVISION TOXICOLOGY LABORATORY CHAIN-OF-CUSTODY

DATE (MM/DD/YY) 05/05/23	ESD TEST NAME	XTEST	NAME OF SAMPLER	GARLAND	/WILLIAMS	PAKC	CHAIN-OF-CUSTODY NO	1102
SAMPLE NAME	OUTFALL NUMBER	SAMPLING TIME	SAMPLE TYPE *	NO. OF CONTAINERS	TOTAL VOLUME	TEMP (°C)	# 7009 RE	MARKS # 4102 C/2
TOX TEST	200	07/0	C	/	~ 17LITERS	He	11.6	<0.05
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En Provinci								
			A.	Le st			Γ,	
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					(2.3			~
3 - 2								
THERMOMETER NO.								
SAMPLES RELINQUISHED BY	& Juli	and			DAT	5/5/23	TIME 08	2 <u></u> <u> </u>
	rijia ku				DAT	5/5/23	TIME O 8	 ⊃ 2 □ PM

UCN-18631 (3 3-92)

* GRAB (G), 24 HR. COMPOSITE (C), OR OTHER (O; DESCRIBE)

ENVIRONMENTAL SCIENCES DIVISION TOXICOLOGY LABORATORY CHAIN-OF-CUSTODY

DATE (MM/DD/YY), 05/08/23	ESD TEST NAME	XTEST	NAME OF SAMPLER	GARLANDI	WILLIAMS	FAIR	CHAIN-OF-CUSTODY NO. 031103
SAMPLE NAME	OUTFALL NUMBER	SAMPLING TIME	SAMPLE TYPE *	NO. OF CONTAINERS	TOTAL VOLUME	FRIG TEMP (°C)	47009 REMARKS 44102 TEMP C/2
TOXTEST	200	0715	C	1	2.16.51	40	14.7 <0.05
				la que			
		4		2 C			
				1	8/		
					ES ES		
THERMOMETER NO.							<u> </u>
SAMPLES RELINQUISHED BY	Jaila	nd				ATE 5/8/2	3 ^{TIME} 0823 [™]
	ia Kn			ŝ	D	ATE 05/08/3	TIME
UCN-18631 (3 3-92)		* 0	DAR (C) 24 HD	COMPOSITE (C) OR	OTHER (O. DESCRIBE)		

* GRAB (G), 24 HR. COMPOSITE (C), OR OTHER (O; DESCRIBE)

TOXICITY TEST LOGSHEETS

	DJECT N							BOOK NO
		Toxi	icity Test	Informati	on Sheet			
Sponsor:	412	Site/Tre	eatment:	OF2	00	Test nu	mber:	2921
Test begin date	e (Dav (l)							
10St Degin date	(Day 0)	act la			cot auratio	/	Tem	plate number
05/03/2	23	05/10	12025		_ · □ hours	🛛 days	□ NA	B 33
Test V Organism:	Z Ceriodaph <u>I</u> Date: <u>5131</u>		123	□ Fathead m Hatch date:	innow		Other:	
Test period		Test purpos	ry		iminary lytical		est type Effluen Receive Substar	ed waters
Treatment desc				1.00		-		
		cription*			Treatment	t Descripti	on*	Type**
1=	DMW 25	10	⊠C ⊡Ţ		50%	6 07-201	0	DCDT
2 =	12.5% 1	07-200	D C 🛛 T	5 =	.75	2 0F20	0	DCUT
3 =	21% 2	0F200		6 =	100	20 0F-20	R	DCUT
				-				
☑ 25% 1	Type: pplicable Dilute Miner	□ al Water (DM	Other (desc		2			
Dilution Water □ Not ap 25% I Source of Test	Type: pplicable Dilute Miner Organisms	□ al Water (DM	Other (desc W) + Trace	ribe): Metals	2			
Dilution Water □ Not ap 25% I Source of Test	Type: pplicable Dilute Miner Organisms cultures: Boa	□ al Water (DM	Other (desc ₩) + Trace	ribe): Metals 4762	2	ber: <u>94</u> 7		
Dilution Water □ Not ap 25% I Source of Test	Type: oplicable Dilute Miner Organisms cultures: Boa	□ al Water (DM : rd numbers: [Other (desc ₩) + Trace	ribe): Metals 4762	2			
Dilution Water □ Not ap 25% I Source of Test √ ESD o □ Vendo	Type: oplicable Dilute Miner Organisms cultures: Boa or: dates: oplicable	□ al Water (DM : rd numbers: [Other (desc (W) + Trace \square NA \square \square Other 33285 43287	ribe): 9 Metals <u>4162</u> (describe): _ Date: Date:	2	ber: <u>94</u>	03110	
Dilution Water □ Not ap √ 25% I Source of Test √ ESD o □ Vendo Water delivery	Type: oplicable Dilute Miner Organisms cultures: Boa or: dates: oplicable	al Water (DM : rd numbers: [Sample ID: _ Sample ID: _ Sample ID: _	Other (desc: W) + Trace \square NA \square \square Other 33286 43287 33288	ribe): Metals (describe): (describe): Date: Date:	Batch num 25/03/23 5/05/23 05/08/	ber: <u>94</u>	03110 031102 031102	
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Dilution Water □ Not ap √ 25% I Source of Test √ ESD o □ Vendo Water delivery □ Not ap	Type: oplicable Dilute Miner Organisms cultures: Boa or: dates: oplicable Record o	al Water (DM al	Other (desc: W) + Trace \square NA \square \square Other 33286 43287 33288	ribe): Metals (describe): (describe): Date: Date:	Batch num 25/03/23 5/05/23 05/08/	ber: <u>94</u>	03110 031102 031102	3
Dilution Water □ Not ap √2 25% I Source of Test √2 ESD o □ Vendo Water delivery □ Not ap Date	Type: oplicable Dilute Miner Organisms cultures: Boa or: dates: oplicable <u>Record o</u> Descript	al Water (DM al	Other (desc: W) + Trace \square NA \square \square Other 33286 43287 33288	ribe): Metals (describe): (describe): Date: Date:	Batch num 25/03/23 5/05/23 05/08/	ber: <u>94</u>	03110 031102 031102	3 Initial
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						CIIICO	NIC Dal	ly mater.	recuing	LUg			· v	
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Sponsor	: <u> </u>	l'est site/tre	atment:	DFZ	00	Begin Dat	te: 05/0	3/2023 End	d Date: _ 0	5/10/2023	E Test Num	iber: 2	985	
а. к ,					Tee	ding Info	mation		[· ·	
		Tempe Inform		(Fe	ood codes:	YCT = yea	ast-cerophyl		Test I	nitiation, V	Vater Chang	ge, or Test		
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Test day	Date	Chamber	Chamber	Food	Food Prep	Volume	Confirm	Feed Time	Start Time	End Time	Sample ID	Control Water Batch	An	alyte
uay		(C)	(C)	Туре	Date	(µL)	cell density		Time			Number		
D	05/03/23	am	am	YUT	3121123	abi	ZYes.	am	1803	1900	33286	947	61	с <u>л</u> .
Day 0	PZ.	26.0 pm	.25.0 pm	R	513/23	86	3.50E7	<u>1830</u> pm				,-, ,	N	A
Day 1	05/04/23		am 25.0 pm	YCT	3/21/23	100	. v∕ŹYes	am 1650 pm	1/20	1725		947		•
Day I	PK	· · ·		R	5/3/23	9)	3.30Eto]		1630	1/25		(-7)	$ \rightarrow $	
Day 2	05/05/23	am 25.7 pm	25:5 pm	YLT	3/21/23	. 100	3.33E+07	am 1.525 pm	1510	1600	33287	947		
	PIC	25.7 am	•	R YUT	5/3/23 3/21/23	90	v Yes	1625 am						
Day 3	05/06/23	pm	pm	R		91	2.28Et07	pm	1610	1700.		94-7		
	РК	25.7 am	25.5 am	XCT	5/3/23	. (00	ZYes	1505 am	·			· · ·		
Day 4	05107/23 plc	pm	pm	R.	513/23	92	3.25E+07	pm	1450	1600	\bigvee	950		
	id al ai	25.8 am	25325.3 PK	YCT	3/21/23	001	Yes	1535 am	11-11-		22 - 40		•	
Day 5	Pla	n nm	pm	R	513/23	9z	3.25E+07	pm	1515	1635	33288	950		
Day 6	05/09/23	25.6 am		YUT	3/21/23	100	VZYes	<u>1518</u> am pm	1504	1630		950		
Dayo	PIK	pm	pm	R	5/3/23	90	3.345+0							
Day 7	05/10/23	>5.3 am	<u>25.0</u> am pm				DYes-	am	1125	1150				/
	PIK			~							<i>\ </i>	X/////////////////////////////////////	~	
Notes:							۰.			· ·				
Environn	nental Science	s Division						·				Rev. 0)3 2020-	-06-0

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PROJECT NAME

221

NOTEBOOK NO.

	т	Project:	VI2 Ceri	iodaphnia Chro	onic Daily Surv hemical:	vival & Reprodu ひ - 20で	Iction Log	umber: 29.	10 F
	r T	roject:							85
	U.	(05 03 2023 Codes: (-) Alive and I	No Reproduction; ((N) Alive and Repr	oduction; (xN) Dear	d and Reproduction	(M) Male r	A-Spic
ľ	Test	Treatment	Day: 1 PK	2 PK	3 PK-	4 PK		6 34	
	Chamber	Number	Date: 05/04/23	1 05105123	05/06/23	05/07/23		05/09/22	7 PE C5/10/23
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mental Sciences Division

	73
Toxicity Test Information Sheet	in the second
DP	- Carlor
Test begin date (Day 0) Test end date Test duration Template number	
05/03/2023 05/10/2023 7 1 hours p days pNA 1	PK
	- 10 the W.
Test Ceriodaphnia dubia Isolated from	5/02/23
Organism: <u>Isolated from:</u> Hatch date: <u>05/04/2023</u> Notes:	Low
Time: Delivery date: Dt/02/2023	in the second
VI Chronic VI Regulatory I Preliminary I Effluent	
□ Acute □ Investigative □ Analytical □ Received waters	
□ Re-test □ Substance	
Treatment descriptions: Number Treatment Description* Type** Number Treatment Description* Type**	الم المسلك . المسلك .
NumberTreatment Description*Type**NumberTreatment Description*Type** $1 =$ DMW 25% $\square C \square T$ $4 =$ 5% $\square C \square T$	
$2 = \frac{12.5\%}{12.5\%} \square C \square T \qquad 5 = \frac{3.5\%}{75\%} \square C \square T$	
$3 = 25\% \qquad \Box C QT \qquad 6 = 100\% \qquad \Box C QT$	
*If DMW, include Batch number **C = Control, T= Treatment	
II DIVIV, Mondo Buck Barre	
Dilution Water Type:	Life best store
□ Not applicable □ Other (describe):	بالاسب
\square 25% Dilute Mineral Water (DMW) + Trace Metals Batch number: <u>947, 95</u> 0	
Source of Test Organisms:	
\Box ESD cultures: Board numbers: \Box NA \Box	
Vendor: ABS D Other (describe):	
Water delivery dates:	
□ Not applicable Sample ID: 33286 Date: \$5(03/23 COC #: 07/10]	
Sample ID: 3728 Date: 05/05/23 COC #: 571102	
Sample ID: 33288 Date: 05/08/23 COC #: 031103	
Record of Deviations from Method and/or Test Non-Conformities	
	101
Date Description Initial	
Date Description Initial 05/10/23 NA PK	
Date Description Initial 05/10/23 NA PK. Quality Assurance (QA) Record PK.	
Date Description Initial 05/10/23 NA PK. Quality Assurance (QA) Record	
Date Description Initial 05/10/23 NA PK. Quality Assurance (QA) Record Initial Date Procedure Name Initial Date Test run by: Peijia Km PK. 05/10/23 Data sheets QA: NONE NONE NONE	
Date Description Initial 05/10/23 NA PK. Quality Assurance (QA) Record Initial Date Procedure Name Initial Date Test run by: Peijia Km PK. 05/10/23	

the state

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CHRONIC Daily Water/Feeding Log

Daily	y Test Info	Tempe Inform Therm. #:_			ood codes: S= Selene	estrum, B=]	r mation ast-cerophyl Brine shrim e = 3.0 to 3.	p)	Test Ir	ge, or Test	Sample Info		
Test day	Date	Env. Chamber (C)	Test Chamber (C)	Food Type	Food Prep Date	Volume (µL)	Confirm a lgae count	Feed Time	Start Time	End Time	Sample ID	Control Water Batch Number	Analyte
Day 0	05/03/23 PEL/NUN	am 264 pm	am 153 pm	B	05/202/22	195	□NA ØYes	am 1752pm	1630	1742	33,286	94-7	NA
Day 1	05/04/23 P/C	26.2 am 26.3 pm	25.7 am 25.9 pm	B B	05/03/04	, 79 94	□NA ØYes	1030 am 1625 pm	1325	1420	J	947	
Day 2	05/05/23 PK	26.2 am 26.4 pm	25.7 am 25-9 pm	вВ	0510423	189	□NA IIIYes	0830 am 1455 pm	1200	1255	33287	947	
Day 3	05/06/23 DE	<u>26.6</u> am <u>25.3</u> pm	26.° am 24.3 pm	B B	05/05/23		□NA ØYes	0910 am 1604 pm	1450	itro		947	
Day 4	05/07/23 PK	<u>26.1</u> am <u>27.9</u> pm	<u>ンド、Y</u> am <u>マゲ-ゲ</u> pm	B B	05(06/23 07/06/23	1.1	□NA D∕Yes	<u>0950 am</u> 1621 pm	1240	1340	1	950	
Day 5	07/08/23 PK	<u>26.0</u> am	<u>25.5</u> am 25.7 pm	B B	03/07/23 05/07/23		□NA ØYes	0845 am 1505 pm	1250	1335	37288.	950	
Day 6	05/09/23 PK	26.0 am 26.0 pm	<u>25.5</u> am 25.5 pm	B	05/08/23	91 59	□NA ØYes	0 <u>935</u> am 1700pm	1210	1310	V	950	
Day 7	05/10/23 PK	26.70 am pm	25.5 am pm						1500	1630-			NA

Sponsor: 12 Test site/treatment: 0F200 Begin Date: 05/03/2023 End Date: 05/10/2023 Test Number: 1692

Notes:

Environmental Sciences Division

Rev. 01 2019-05-28

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			· Fa	thead Minnow	v Chronic Dai	y Survival Lo	g		N
Sponsor:	YIZ		_ Test site/ch	emical:	OFDO		Test Numbe	r: 1692	PK 05/02
Begin Date:	051	03/2023	End Date:	05/10/2023					
0			Fg = Fungus; K =			Sk = Sick; SM = S	mall; SOR = Sipho	oned and returned;	W = Wounded
Treatment	~ ••		Day 1	Day 2	Day 3	Day 4	Day 5	Day 6.	Day 7
Number	Replicate Number	Position Number	Date	Date	Date	Date	Date PK	Date PK	Date PK
and Desc.	Itumber	Tumber	05104723 PK	05/05/23 PK	05106/23 AC	1)5/07/23 PK	05/08/23	05/09/23	05/10/23
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25% pm	· 2	24	10	10	10	. 10	10	10	10
2310 Marrie	3	11	,0	10	10	10	10	10	10
a. 4	4	14	10	10	10	10	10	10	10 .
2:	1	13	10	10	10	10	10	10	QIU OPE
12.5%	2	10	10	10	10	10	10	10	10
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3: 25Z -	1	21	10	10 .	10	PIC 0100	10	10	10
	2	12 .	10	0	pt 10 @	10	10	20	10
0F200	3	19	9 ID	9	8.10	8	8	8	8
	4	3	10	ID	i0 ·	10	10	10	10
4:	1	7	10	10	10 .	9' IP	9	9	9
4: 50%	2	20	10	(0	10 ISK	10	10	10	10
07200	3	1	10	10	10	10	10 .	10	10
	4	22	10 0 15 15 PM	10	10	10	10	10	10
5:	- 1	23	10	10	10	10	10	10	10
5: 75%	2	4	10	10	PK 609 1D	9	9	9	9
07-200	• 3	17	10	10.	10	10	10	10	10
	4	2	10	10	10	10	10 .	10	10
6:	1	5	10	ru zem	10 24M	10 2511	10° 25M	10 25M	10 25/19
100%	2	9	(0)	ų O	10	10	10	10	10
07-200	3	6	10	10	10	10	12.	10	10
	4	R	10	10	10	10	D	10	10

Random Assignment of Test Chambers V12 0F200 Test site/chemical: **Project:** Test Number: Starting Position Number (on Table of Random Numbers): 29 pk-Sample ID/Treatment Assigned Numbers Replicate Position 05/02/23 50%, 0F200 - 3 75% 07-200 -4 218 0F200 3--4 -2 75% DF-200 6. 100% UF200 -1 100% UF200 -3 Joz of-200 4--1 100% DF200 ~4 6-100% 0F200 -2 12.5% 0T-200 -2 25% DMW N 1. -3 25% OF200 -2 125% JF200 -1 25% PMW -4 1-12.5% 0F-200 2. -4 12.5% OF200 2-- 3 75% OF-200 -3 25% DMW 1--1 25% 0F200 3--3 56% 0F200 4-2 21% 0F200 3-70-50% 0F200 4--4 75% 0F200 25% DMW

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ponsor:	Y12		Test nu	mber: 1692	
Test site/chemi		õ	Balance		
Test Start Date:	V			id Date: 05/10/2	
				ying Date/time: 05	
	<u>9</u> 910	29,100		, <u> </u>	1
		Pan W	t. (mg)	Pan + Larvae (mg)	Number
Treatment	Replicate	Date:_1 Balance	05/09/23 check:	Date: 05/11/23 Balance check:	Surviving
nitial	1	15	.1175	16.4060	10
	2		10915		/
	3	15.	0130		
	4	15.	0090		
•	1	15.	0785	23.2285	10
	2	15.	.0645	22.8365	10
5% DMW	3	15	.2950	22.8305	10
	4	15.	2245	22.7680	10
	1	15	1305	23.7075	10
2.5%	2	14	1785	21.8920	10
0T-200 -	3	15.	0895	PK 22 21.9525	9
0-200	4	15.	2905	23.1345	10
57	. 1	115	2900	22.7785	10
25%	2	_ 15.	2875	23.9815	10
05-200	3	115.2	3000	22,3025	8
	4	15.	3370	23.0570	10
5.9	1	115.	33307	22.5860	9
50%	2	15.2	855	23.7705	10
0F-200	3		1690	24.4465	(0)
	4		3905	22.3495	10
in the W	1		3705	22.8755	10
75%	2	15.	3250	22.3335	9
07-200	3	15.	1295	22.3390	10
	4		3605	23.5665	01
	1		3685	22.4470	10
100% 0F200	2		210	22.2235	10
07-200	3		780	23,6150	10
	4		3780	23.1690	10

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and the second

		Random Assignment of Larvae to Test Chambers												
	Proje	ect:	YIT	k. 1	KU.	Ref Test site/che	0					Test Number:	1692; 169	93
					•	(on Table of Ra								-
PK		ing i	112		ATTANCAS (14			C13)			KC	1		
				gned		Sample	Replicate	Assigned Numbers			Sample	Replicate		
USIC	2/23	V	Nun	ibers		ID/Treatment	1	7			20	ID/Treatment 1.		
		A	20	49	73		14,221 V	N	25	49	73		2,441 1	-
		2		30	74	25% DINW	3,82 V	2	26	30	\$4	25% pmw	18,352 1	-
	r.	R	27	51	75			3	27	31	75		DIF V	-
		X	28	52	76		39; 3/4 J	4	28	32	76	-	1,324 V	-
		5	29	53	·77	2.	5,231 V	X	29	53	X7		3,131 √	·
	·	8	30	54	78	12.5% JF200	21,442 V	6	30	34	78	0.2591L	22,282 V	-
		X	\$1	55	79		27,293 1	X	31	55	79	0.25g/L KCJ	9,233 1	-
		8	32	56	80		1,374 V	8	32	36	80		7,214	-
		Q	33	57	81	3.	18,241 J	X	33	57	81	3.	16,171 V	
		10	34	58	82	25% 0F200	41,482 V	N	34	38	82	0.5g/L	11,142 1	
		N	35	59	83	0-200	19,313 J	N	35	59	83	KU	20,303 V	
		12	36	6Q	84		15,204 1	12	36	60	84		15,424 √	
		R	JE	61	85	4. 50%	25,301 V	R	37	61	85	4.	34,471 J	
	-	14	38	82	86	4. 50% OF200	28,352 V	M	38	62	86	1.09/1	5,821	
		Ìs	38	63	87	01-00	16,45 V	[.] 15	32	63	88	KU	19,463 V	
	, i c	16	40	64	38		7,334 √	16	40	64	88		26,404 V	
	*	17	41	05	89	5.	4,321 V	N	41	65	89	5.	33, BB9 J	
		18	42	66	90	75% 0F-200	2,132 1	18	42	66	30	1.25g/L	6,272 1	
	. ·	N	43	78	91	07-200	6,263 V	19	43	76	À	1.25g/L KCI	4,443 J	
		20	44	68	92	<i></i>	12,394 1	20	À	86	92		37,484 V	
		21	45	82	33	6. 100%	9,421 V	21	45	69	93	6.	31,431 1	-
		22	46	TQ	94	100%	38,472. √	22	46	70	94	1.50g/L	10,122 Y	-
		23	4	71	26	0-2010	11,173 V.	23	47	X	95	KC	36,383 √	-
		\$4	48	72	96	*	43,464 V	24	48	72	96		24,254 1	
	E	nviro	onme	ntal S	Scien	ces Division	terrange and the subscription of					Rev. 0	1 2019-04-2:	5 —

Fathead Minnow Order & Shipment Log

Ordering Information:

Date Ordered	Test #(s)	Vendor	Quantity ordered	Description (larval age, etc.)	Expected delivery	Ordered by	Comments
05/01/23	1692 + 1693	ABS	600	iday old on currival	05/02/23	AMF	

Delivery Information:

Larva source	Approx. number received	Date/time received	Received by (Initials)		
ABS	660	05/02/23 1300	РК		

Monitoring Interval	Hour									
	0	1	2	3	4	5	6	7		
Temperature (°C)	13.5	22.5		23.4						
Time	1305	14:25		1625	1		8			
Thermometer ID	0019	DD19	а 2	1019	*					
Initials	pk	PK.		PK						

Comments (e.g. condition of larvae received):

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px 2/23

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